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SUMMARY

An examination of 38 faecal samples using Polymerase chain reaction (PCR) from buffalo calves with and without diarrhoea for detection of Salmonella genes, revealed the detection of a total of 13 genus specific (16SrRNA) genes which accounts for 11 (28.95%) positive samples. Despite low Salmonella detection rate, the number may still contribute to occurrence of diarrhoea in these animals. It is therefore suggested that larger samples should be collected for a wider screening.

Keywords: Buffalo Calves, Diarrhoegenic, Non-diarrhoegenic, Faeces, PCR, Salmonella

In bovines, Salmonella infections comprise the second most economically important bacterial disease affecting the gastrointestinal system following E. coli infections. (Ekperigin and Nagaraja, 1998). Despite the fact that most bacterial pathogens can be easily cultured, there are a number of problems associated with attempts to detect and quantify them. Consequently, rapid, accurate and culture-independent alternatives are being investigated to enable detection of pathogens in diarrhoea cases (Alexandrino et al., 2004). Different molecular typing methods based on variations in genetic makeup have been now used in complement with traditional typing methods for fingerprinting of Salmonella serotypes. The present study was aimed at molecular screening of faecal samples from buffalo calves for Salmonella spp. (Rakesh et al., 2009).

Faecal samples from 38 buffalo calves were collected from buffalo farm in Lala Lajpat Rai, University of Veterinary and Animal Sciences (LUVAS), Hisar. Buffalo calves were grouped on the basis of age *viz*. Group1 (day1 to day2) Group2 (Day 3 to 3 months) and Group3 (4 to 6 months). Twenty of 38 calves were diarrhoeic and 18 were healthy clinically.

One gram of each faecal sample was processed for extraction of DNA to detect various genes in Polymerase chain reaction (PCR) assays (Invitrogen, 2018).

PCR was carried out using primer pair Sal-F (5' -TGTTGTGGTTAATAAC CGCA-3') and Sal-R (5'-CACAATCCATCTCTGGA-3') for amplifying (16s rRNA) coding gene complicon size 544 bp supplied by Thermo[®] (USA). Thermal cycle comprised of initial denatuvation at 98 °C for 3 min followed by 40 cycle of denaturation (98 °C for 1 min), annealing (51.4 °C for 45 sec.); extension (72 °C for 2 min) and final extension at 72 °C for 10 min.

The PCR assays were carried out in 25 μ l reaction volume containing 1U of Taq polymerase, 200 μ mol of each dNTP and 2.5 μ l of 10× PCR buffer. Reactions were performed in a GeneAmp[®] PCR System 2400 Thermal Cycler (Applied Biosystems, California) and FlexiGene[®] thermal cycler (Techne Inc., New Jersey).

An examination of 38 faecal samples from buffalo calves revealed that 11 were positive for salmonella using genus specific genes. Chi-square analysis of Salmonella positivity was assessed for effect of presence of diarrhoea, age and sex and was found non-significant (P>0.05).

The present study was conducted to detect the presence of Salmonella infection from diarrhoeic and nondiarrhoeic buffalo calves using polymerase chain reaction and recorded the prevalence of 28.95 (11/38).

Similar observations were recorded by Fahmy et al. (2017) after examining faeces of calves from feedlot farms in Egypt but the percentage of Salmonella isolated from diarrhoeic calves was 7%. The findings of El-Shehedi et al. (2015) who reported the prevalence of Salmonella infection after examining faecal samples from diarrhoeic calves in Egypt was also lower compared with the findings in the present study. Various workers reported varying degree of prevalence of Salmonella from diarrhoeic faeces viz. 4.0% (Haggag and Khaleel, 2002); 4.09% (Younis et al., 2009); 1.8%, (Garcia et al., 2000); 2.0% (Acha et al., 2004) and 1.56% (Osama et al., 2011). The variations in prevalence of Salmonella among apparently healthy and diarrheic calves reported in different countries reflect the effect of wide range of different management risk factors (Vanselow et al., 2007 & Jones, 2011). Wani et al., 2013, however reported

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Escherichia coli but no *Salmonella* from the faecal samples collected from buffalos' calves of Kashmir region in India. This was further supported by reports from other parts of India and Mozambique according to Hussain and Saikia (2000) and Acha *et al.* (2004), respectively who in their separate studies recorded negative outcomes. Despite low *Salmonella* detection rate, the number may still contribute to occurrence of diarrhoea in these animals. It is therefore suggested that larger samples should be collected for a wider screening.

REFERENCES

- Achá, S.J., Kuhn, I., Jonsson, P., Mbazima, G., Katouli, M. and Mollby, R.S. (2004). Studies on calf diarrhea in Mozambique: prevalence of bacterial pathogens. *Acta Vet. Scand.* 45: 27-36.
- Alexandrino, M., Grohmann, E. and Szewzyk, U. (2004). Optimization of PCR based methods for rapid detection of *Campylobacter jejuni*, *Campylobactercoli* and *Yersinia enterocolitica* serovar 0:3 in wastewater samples. *Water Research*. 38: 1340-1356.
- Ekperigin, H.E. and Nagaraja, K.V. (1998). Microbial food borne pathogens. Salmonella. Vet. Clin. North Am. Food Anim. Pract. 14: 17-29.
- El-Shehedi, M.A., Mostafa M.E. and Hanan, E.N. (2015). Some bacteriological studies on *Salmonella* infection in buffalo calves using PCR technology. *Eur. Scientific J.* 11(36): 181-192.
- Fahmy, M., Yousef, H.M. and Soliman, Y.A. (2017). Detection of Salmonellae in the Feces of Feedlot Calves Farms in Egypt. *Inter. J. Microbiol. Res.* 8(1): 01-08.
- Garcia, A., Ruiz-Santa-Quiteria, J.A., Orden, J.A., Cid, D., Sanz, R. and Gómez-Bautista, M. (2000). Rotavirus and concurrent infections with other enteropathogens in neonatal diarrheicdairy calves in Spain. Comp. Immunol. Microbiol.

Infect. Dis. 23: 175-183.

- Haggag, Y.N. and Khaleel, S.A. (2002). Public health importance of certain bacteria isolated from calves and small ruminants. In: The Proceedings of 2nd Vet. Cong. Fac. Vet. Med., Minufiya University, Egypt. 2(1): 173-184.
- Hussain I. and Saikia G.K. (2000). Isolation and characterization of bacteria from diarrhoeic calves, *Indian J. Comp. Microbiol. Immunol. Infect. Dis.* 21(1): 125–127.
- Invitrogen (2018). Bacterial DNA extraction protocol, using PureLink Genomic DNA purification kit Invitrogen U.S.A.
- Jones, F.T. (2011). A review of practical Salmonella control measures in animal feed. *The J. Appl. Poult. Res.* **20**: 102-113.
- Osama, N.M., Adel, F.F., Amani, F.A. and Rania, F.F. (2011). Fecal shedding of non-typhoidal *Salmonella* species in dairy cattle and their attendants in Alexandria suburbs. *J. Am. Sci.* **7(9)**: 623-631.
- Rakesh, K., Surendran, P.K. and Nirmala, T. (2009). Biochemical and Molecular investigations on Salmonella serovars from seafood. Ph.D. dissertation submitted to Cochin University of Science and Technology, India.
- Vanselow, B.A., Hornitzky, M.A., Walker, K.H., Eamens, G.J., Bailey, G.D., Gill, P.A., Coates, K., Corney, B., Cronin, J.P. and Renilson, S. (2007). Salmonella and on-farm risk factors in healthy slaughterage cattle and sheep in eastern Australia. *Aus. Vet. J.* 85: 498-502.
- Wani, S.A., Hussain, I., Beg, S.A., Rather, M.A., Kabli, Z.A., Mir, M.A. and Nishikawa, Y. (2013). Diarrhoeagenic *Escherichia coli* and salmonellae in calves and lambs in Kashmir absence, prevalence and antibiogram. *Rev. Sci. Tech.* 32(3): 833-840.
- Younis, E.E., Ahmed, A.M., El-Khodery, S.A., Osman, S.A. and El-Naker, Y.F. (2009). Molecular screening and risk factors of enterotoxigenic *Escherichia coli* and *Salmonella spp*. in diarrheic neonatal calves in Egypt. *Res. Vet. Sci.* 87: 373-379.