PHYSICAL AND ANALYTICAL CHARACTERISTICS OF UTERINE DISCHARGE IN SUBCLINICAL AND CLINICAL ENDOMETRITIC BUFFALOES

 THANGAMANI. A*, MANDA SRINIVAS¹, M. MUTHA RAO², MAKKENA SREENU³ and K. ASWANI KUMAR⁴ Department of Veterinary Gynaecology and Obstetrics, Veterinary College and Research Institute,
 Tirunelveli-627358 (Tamil Nadu), ¹Department of Veterinary Gynaecology and Obstetrics, Veterinary College and Research Institute, Salem-636112, TANVASU, India, ²Livestock Research Station, Guntur-522018 (A.P.)
 ³Department of Veterinary Surgery and Radiology, College of Veterinary Science, Garividi, Vizianagarem (A.P.)
 ⁴Department of Veterinary Biochemistry, NTR College of Veterinary Science, Gannavaram-521102 (A.P.), India

Received: 26.11.2022; Accepted: 28.01.2023

ABSTRACT

The present study was undertaken to estimate the physical (Turbidity, Specific Gravity and Electric Resistance), analytical characteristics (Sodium, Potassium and Chloride) and enzyme assay (Leucocyte esterase, catalase and SOD) of uterine discharge in subclinical (SCE) and clinical endometritic (CE) in comparison with normal buffaloes without endometritis. Physical parameters revealed that mean concentrations of turbidity, specific gravity (SG) and electrical resistance (ER) was significantly (p<0.05) higher in CE as compared to SCE and normal buffaloes. Analytical parameters revealed that mean electrolyte concentrations (potassium and chloride) were significantly higher (p) in CE buffaloes as compared to SCE affected and normal healthy buffaloes. Also, Enzymes like leucocyte esterase and catalase showed increasing trend however SOD showed decreasing trend with the severity of infection in uterine discharge of Normal, SCE and CE buffaloes. In conclusion, estimation of physical and analytical parameters of uterine discharge could be a novel non-invasive method for diagnosis of degree of endometritis in affected buffaloes.

Keywords: Buffalo, Electrical resistance, Enzymes, Endometritis, Specific gravity, Turbidity, uterine discharge

How to cite: Thangamani, A., Srinivas, M., Rao, M.M., Sreenu, M. and Kumar, K.A. (2023). Physical and analytical characteristics of uterine discharge in subclinical and clinical endometritic buffaloes. *Haryana Vet.* **62**(SI-2): 105-108.

The key for optimum fertility in dairy cow and buffalo herds is the healthy or ambient uterine environment since any disturbances of endometrium affected the normal reproductive functions and culminated to infertility or subfertility in terms of repeat breeding (Jabbour et al., 2009; Sheldon et al., 2009). Postpartum uterine infection has been associated with the delayed uterine involution followed with increased service period and in turn increased calving intervals culminating in economic losses (Gahlot, 2016). Subclinical endometritis is an important cause of repeat breeding and usually remains undiagnosed (Dutt et al., 2017). Endometrial cytology is considered as a reference method for cytological diagnosis of subclinical endometritis, because of quality of sample obtained by either low volume uterine flushing or cytobrush with great repeatability of the test (Kasimanickam et al., 2005). Uterine discharge molecules could be used to diagnose the SCE in buffaloes (Gahlot et al., 2017). However, no published data was available on physical (turbidity, specific gravity and electrical resistance) and analytical parameters like electrolyte concentration (sodium, potassium, chloride) and enzymes (leucocyte esterase, catalase and superoxide dismutase) pertaining to subclinical and clinical endometritis in buffaloes. In this context, the present study was designed to estimate the changes in physicaland analytical parameters of uterine discharge of SCE, CE affected in comparison with normal healthy buffaloes without endometritis.

MATERIALS AND METHODS

The lactating postpartum buffaloes with history of failure of conception even after three consecutive artificial inseminations (repeat breeder) presented to the Gynaecology Unit, NTR College of Veterinary Science, Gannavaram and buffaloes maintained at informal small dairy units located in and around Gannavaram, Krishna District, Andhra Pradesh were screened. The uterine discharge was collected by uterine flushing technique from all the selected buffaloes at the time of estrus as described by Nehru et al. (2019). Clear uterine discharge with less than 5% of PMN cells and clear uterine discharge with more than 5 percentage PMN on cytology considered as normal and SCE affected buffaloes, respectively. However, uterine discharges with flakes of pusand abundant PMN cells on cytology were considered as CE as described by Sheldon et al. (2009) and Gahlot et al. (2016). An attempt was made to estimate the physical (Turbidity, Specific gravity and Electrical resistance) and analytical characteristics (electrolytes, enzymes) of uterine discharge of normal (n=10) and endometritis (subclinical (n=10) and clinical (n=10)) affected buffaloes on the day of estrus when presented for treatment.

Physical characteristics of uterine discharge:

Turbidity: Turbidity of uterine discharge sample was measured by using digital Nephelo-Turbidity meter and expressed in Nephelometric turbidity unit (NTU).

^{*}Corresponding author: thangamtamil19@gmail.com

Specific gravity (SG): Specific gravity of uterine discharge sample was measured by using digital specific gravity meter and expressed in g/cm^3 .

Electrical resistance: Electrical resistance of uterine discharge sample was measured by using electrical conductivity meter as per procedure described by Ahmed *et al.* (2017) and expressed in Ohm (Ω).

Analytical characteristics of uterine discharge:

Electrolytes concentration: Electrolytes concentration of uterine discharge sample were measured by using Elyte kit (Tulip diagnostics Pvt. Ltd.,) and expressed in mmol/l.

Enzymes concentration:

Leucocyte esterase enzyme: Leucocyte esterase enzyme concentration of uterine discharge sample was assessed by using Leucocyte esterase ELISA Kit and expressed in ng/ml.

Catalase enzyme: Catalase enzyme concentration of uterine discharge sample was assessed as procedure described by Aebi (1983) and enzyme activity was calculated as mM H_2O_2 utilized/min/mg protein, using a molar extinction coefficient of 43.6 M-1 cm-1.

Superoxide dismutase (SOD) enzyme: SOD enzyme concentration was estimated as procedure described by Madesh and Balasubramanian (1998).

RESULTS AND DISCUSSION

The mean level turbidity was significantly (P ≤ 0.01) higher in CE affected buffaloes (0.55±0.19) than SCE (0.14±0.31) affected and normal healthy buffaloes (0.05±0.00). However, the mean levels of turbidity were non-significantly (P>0.05) higher in the uterine discharge of SCE buffaloes (0.14±0.31) than normal (0.05±0.00) healthy buffaloes (Table 1). The mean levels of turbidity were significantly higher in CE affected buffaloes might be due to the presence of higher bacterial load and excessive levels of metabolites in the uterine lumen of buffaloes affected with different grades of endometritis (Bondurant, 1999 and Miller *et al.*, 2007). It was inferred that turbidity level in uterine discharge was increased in accordance with degree of uterine infection.

The mean specific gravity (SG) was significantly (P \leq 0.05) higher in CE affected buffaloes (1.10±0.00) than SCE (1.06±0.00) affected and normal healthy buffaloes (1.03±0.00). However, the mean SG was non-significantly (P>0.05) higher in the uterine discharge of SCE buffaloes (1.06±0.00) than normal (1.03±0.00) healthy buffaloes (Table 1). The mean levels of SG were significantly higher in clinical endometritis affected buffaloes correlated with presence of extra substances in the uterine discharge *viz.*,

debris, cellular components, inflammatory exudates, bacterial metabolites and probably flakes of pus or purulent material (Machado *et al.*, 2012). It was inferred that SG could help the clinician to differentiate between SCE and CE when used in combination with turbidity assessment of uterine discharge sample.

The mean electrical resistance (ER) was significantly ($P \le 0.01$) higher in CE affected buffaloes (180.09 ± 7.71) than SCE (148.15 ± 8.43 Ù) affected and normal healthy buffaloes (132.06 ± 7.80). However, the mean ER was non-significantly (P > 0.05) higher in the uterine discharge of SCE buffaloes (148.15 ± 8.43) than normal (132.06 ± 7.80) healthy buffaloes (Table 1). The mean electrical resistance(ER) was significantly higher in CE affected buffaloes might be due to the presence excessive inflammatory products in the uterine discharge.

Collectively, the mean concentrations of turbidity, SG and ER was significantly (P<0.05) higher in CE affected buffaloes as compared to SCE affected and normal healthy buffaloes. However, mean concentrations of turbidity, SG and ER were non-significantly (P>0.05) higher in the uterine discharge of SCE buffaloes than normal healthy buffaloes.

The mean concentrations of sodium was significantly $(P \le 0.05)$ lower in CE affected buffaloes (82.22 ± 5.04) than SCE (104.19±3.94) affected and normal healthy buffaloes (118.19±6.27). However, mean concentrations of sodium was non-significantly (P>0.05) lower in the uterine discharge of SCE buffaloes (104.19±3.94) as compared to normal (118.19±6.27) healthy buffaloes. The mean concentrations of potassium was significantly ($P \le 0.05$) higher in CE affected buffaloes (11.59±1.11) than normal healthy buffaloes (7.82 ± 0.56) , while the potassium concentration was moderately higher in SCE (9.10 ± 1.05) affected buffaloes as compared to normal healthy buffaloes (7.82±0.56) and moderately lower as compared to CE affected (11.59±1.11) buffaloes. The mean concentrations of chloride was significantly ($P \le 0.05$) higher in CE affected buffaloes (36.18±3.10) as compared to SCE (27.82 ± 1.36) affected and normal healthy buffaloes (24.06 ± 1.69) . However, the mean concentrations of chloride was non-significantly (P>0.05) higher in the uterine discharge of SCE buffaloes (27.82±1.36) than normal (24.06±1.69) healthy buffaloes (Table 2). Higher concentrations of chloride and potassium in the uterine discharge of the present study reflected the increased concentration of electrolytes present in the lumen of the uterus in relation to degree of uterine infections as a result of endometrial insult in cyclical animal as stated by Igarashi (1954) and Heap (1962).

Table 1. The physical characteristics of uterine discharge (Mean±SE) in normal, subclinical and clinical endometritic buffaloes

Physical characteristics	Normal (n=10)	Subclinical endometritis (n=10)	Clinical endometritis (n=10)	F-value
Turbidity (NTU)	$0.05{\pm}0.00^{\text{b}}$	$0.14{\pm}0.31^{\text{b}}$	0.55±0.19ª	5.667
Specific gravity (g/cm ³)	$1.03{\pm}0.00^{\text{b}}$	$1.06{\pm}0.00^{\text{b}}$	$1.10{\pm}0.00^{\circ}$	28.141
Electrical resistance (Ω)	$132.06 \pm 7.80^{\text{b}}$	148.15±8.43 ^b	$180.09 \pm 7.71^{\circ}$	9.357

Means bearing different superscripts (ab) differ significantly (P≤0.01 or P≤0.05) within a row

 Table 2.
 The electrolyte concentration (Mean±SE) in uterine discharge of normal, subclinical and clinical endometritic buffaloes

Electrolyte concentration	Normal (n=10)	Subclinical endometritis (n=10)	Clinical endometritis (n=10)	F-value
Sodium(mmol/l)	118.19±6.27 ^ª	104.19±3.94ª	82.22±5.04 ^b	12.259
Potassium(mmol/l)	$7.82{\pm}0.56^{\circ}$	$9.10{\pm}1.05^{ab}$	11.59±1.11ª	4.145
Chloride(mmol/l)	24.06±1.69 ^b	27.82±1.36 ^b	36.18±3.10 ^a	8.044

Means bearing different superscripts (ab) differ significantly (P≤0.01 or P≤0.05) within a row

Electrolyte concentration	Normal (n=10)	Subclinical endometritis (n=10)	Clinical endometritis (n=10)	F-value
Leucocyte esterase (ng/ml)	$0.17{\pm}0.01^{\circ}$	$0.24{\pm}0.05^{ab}$	$0.28{\pm}0.07^{a}$	5.951
Catalase (μ mol of H ₂ O ₂ Consumed/min/mg of protein)	52.94±3.74 ^b	63.17 ± 4.88^{ab}	68.89±3.94ª	3.662
Superoxide dismutase (Units/min/mgof protein)	4.65±0.21 ^b	4.10±0.28 ^b	2.61 ± 0.30^{a}	15.032

Means bearing different superscripts (ab) differ significantly ≤ 0.01 or P ≤ 0.05) within a row

On the whole, the mean electrolyte concentrations (potassium and chloride) were significantly altered in CE affected buffaloes as compared to SCE affected and normal healthy buffaloes. However, the mean concentrations of potassium and chloride (mmol/l) were not much altered in the uterine discharge of SCE buffaloes than normal buffaloes.

The mean concentrations of leucocyte esterase enzyme (LE) were significantly (P \leq 0.05) higher in CE affected buffaloes than SCE affected and normal healthy buffaloes, (Table 3). The mean concentrations of catalase enzyme were significantly (P \leq 0.05) higher in CE affected buffaloes than SCE affected and normal buffaloes (Table 3).

The mean concentrations of SOD were significantly ($P \le 0.05$) lower in CE affected buffaloes than normal and SCE affected buffaloes. However, mean concentrations of SOD were not much altered in the uterine discharge of subclinical endometritic buffaloes and normal healthy buffaloes (Table 3). Enzyme concentrations in uterine discharges of normal, SCE and CE affected buffaloes were

increased in concentration especially that of LE (ng/ml) and catalase (μ mol) enzymes, which increased with severity of uterine infection. While, SOD activity was decreased with increase in severity of uterine infection.

The results of present study were comparable with the reports of Gahlot *et al.* (2017) who observed LE concentrations (IU/L) in uterine fluid were nonsignificantly differed between CE buffaloes and normal buffaloes, similarly between SCE and normal buffaloes. Variations in the concentration of LE in the present study might be due to the activity of LE, an enzyme produced from neutrophils and was elevated in inflammatory exudate (Cheong *et al.*, 2012, Couto *et al.*, 2013 and Nazhat *et al.*, 2018). Increased concentration of catalase enzyme in CE culminated to the generation of free radicals, which in turn aggravated the inflammatory reaction resulting in reduction of uterine muscle activity and reduced the uterine muscle tone causing build-up of inflammatory products in the lumen of the uterus, thus increased the severity of uterine inflammation (Li *et al.*, 2010). Further, these changes adversely affected the life of spermatozoa during its transport through the uterus eventually resulting in fertilization failures as opined by Zhong and Zhou (2013). SOD activity decreased with increase in severity of uterine infection in the present study was in conformity with the investigation of Kaya *et al.* (2017) who reported that total antioxidant capacity decreased in endometritis affected animals and inferred that stress related enzymes could be used as biochemical markers for assessment of the severity of endometritis in bovines.

CONCLUSION

Estimation of physical and analytical parameters of uterine discharge could be novel non-invasive method for diagnosis of degree of endometritis in buffaloes as Turbidity, SG, ER, LE and catalase enzymes concentrations increases however SOD activity decreases with the severity of uterine infection.

REFERENCES

- Aebi, H.E. (1983). Catalase. In: Bergemeyer H U (Edn.), Methods of Enzymatic Analysis, Verlag Chemie, Weinheim, Germany, pp. 273-285.
- Ahmed, M.K., Chowdhury, Rahman, M.M., Bhattacharjee, J. and Bhuiyan, M.M.U. (2017). Relationship of electrical resistance of vaginal mucus during oestrus with post-AI pregnancy in cows. *Bangladesh J. Vet. Med.* 15: 113-117.
- Bondurant, R.H. (1999). Inflammation in the bovine female reproductive tract. J. Anim. Sci. 77: 101-110.
- Cheong, S.H., Nydam, D.V., Galvao, K.N., Crosier, B.M., Ricci, A. and Caixeta, L.S. (2012). Use of reagent test trips for diagnosis of endometritis in dairy cows. *Theriogenol.* 77: 858-864.
- Couto, B.G., Vaillancourt, D.H. and Lefebvre, R.C. (2013). Comparison of a leukocyte esterase test with endometrial cytology for diagnosis of subclinical endometritis in postpartum dairy cows. *Theriogenol.* **79**: 103-107.
- Dutt, R., Singh, G., Singh, M., Sharma, M., Dalal, J. and Chandolia, R.K. (2017). Diagnosis of Subclinical Endometritis in Murrah Buffaloes through CytobrushTechnique. *Int. J. Curr. Microbiol. App. Sci.* 6(11): 494-499.
- Gahlot, S.C., Kumar S., Kumaresan, A., Vairamuthu, S., Saraf, K.K. and Mohanty, T.K. (2017). Biochemical analysis of uterine fluid for identification of indicators for subclinical endometritis in the water buffalo (*Bubalus bubalis*). *Reprod. Domest. Anim.* 1: 1-6.
- Gahlot, S.C., Kumar, S, Kumaresan, A, Chand., S, Baithalu., R.K. and Mohanty, T.K. (2016). Efficiency of uterine fluid cytology in the diagnosis of subclinical endometritis in the water buffalo (*Bubalus bubalis*). *Reprod. Domest. Anim.* 2: 1-4.

- Gahlot, S.C. (2016). Biochemical and proteomic analysis of uterine fluid in buffaloes affected with uterine infection (*Bubalus bubalis*). M.V.Sc thesis submitted to National Dairy Research Institute, Karnal.
- Heap, R.B. (1962). Some chemical constituents of uterine washings: a method of analysis with results from various species. *J. Endocrinol.* 24: 367-378.
- Igarashi, M. (1954). Studies on the cyclic changes in human cervical mucus as well as its crystallization phenomenon. *J. Japan. Obst. and Gynae Soc.* **1**: 194-202.
- Jabbour, H.N., Sales, K.J., Catalano, R.D. and Norman, J.E. (2009). Inflammatory pathways in female reproductive health and disease. *Reprod.* 138(6): 903-619.
- Kasimanickam, R., Duffield, T.F., Foster, R.A., Gartley, C.J., Leslie, K.E., Walton, J.S. and Johnson, W.H. (2005). A comparison of the cytobrush and uterine lavage techniques to evaluate endometrial cytology in clinically normal postpartum dairy cows. *Can. Vet. J.* 46: 255-259.
- Kaya, S., Ogun, M., Ozen, H., Kuru, M., Sahin, L., Kukurt, A. and Kacar, C. (2017). The impact of endometritis on specific oxidative stress parameters in cows. J. Hell. Vet. Med. Soc. 68: 231-236.
- Li, D., Liu, Y., Li, Y., Lv, Y., Pei, X. and Guo, D. (2010). Significance of nitric oxide concentration in plasma and uterine secretes with puerperal endometritis in dairy cows. *Vet. Res. Commun.* 34: 315-321.
- Machado, V.S., Knauer, W.A., Bicalho, M.L.S., Oikonomou, G., Gilbert, R.O. and Bicalho, R.C. (2012). A novel diagnostic technique to determine uterine health of Holstein cows at 35 days postpartum. *J. Dairy Sci.* **95**: 1349-1357.
- Madesh, M. and Balasubramanian (1998). Microtiter plate assay for superoxide dismutase using MTT reduction by superoxide. *Ind. J. Biochem. Biophys.* 35: 184-188.
- Miller, A.N., Williams, E.J., Sibley, K., Herath, S. and Sheldon, I.M. (2007). The effects of Arcanobacterium pyogenes on endometrial function in vitro, and on uterine and ovarian function *in vivo*. *Theriogenol.* 68: 972-980.
- Nazhat, S.A., Kitahara, G., Kozuka, N., Mido, S. and Osawa, T. (2018). Associations of periparturient plasma biochemical parameters, endometrial leukocyte esterase and myeloperoxidase, and bacterial detection with clinical and subclinical endometritis in postpartum dairy cows. J. Vet. Med. 80: 302-310.
- Nehru, D.A., Dhaliwal, G.S., Jan, M.H., Cheema, R.S. and Kumar, S. (2019). Clinical efficacy of intrauterine cephapirin benzathine administration on clearance of uterine bacteria and subclinical endometritis in postpartum buffaloes. *Reprod. Domest. Anim.* 54: 317-324.
- Sheldon, I.M., Price, S.B., Cronin, J., Gilbert, R.O. and Gadsby, J.E. (2009). Mechanisms of infertility associated with clinical and subclinical endometritis in high producing dairy cattle. *Reprod. Domest. Anim.* 44: 1-9.
- Zhong, R.Z. and Zhou, D. (2013). Oxidative stress and role of natural plant derived antioxidants in animal reproduction. J. Integr. Agric. 12: 1826-1838.